



	<b>Experiment title:</b> Structural studies of chloroplast F <sub>1</sub> -ATPase	<b>Experiment number:</b> LS-1755
<b>Beamline:</b> ID14-2	<b>Date of experiment:</b> from: 03-11-00 to: 04.11.00	<b>Date of report:</b> 1.2.01
<b>Shifts:</b> 3	<b>Local contact(s):</b> Stéphanie Monaco	<i>Received at ESRF:</i>
<b>Names and affiliations of applicants (* indicates experimentalists):</b> <b>G. Groth*, C. Schnick*</b> <b>Heinrich-Heine-Universität, Biochemie der Pflanzen, D-40225 Düsseldorf, GERMANY</b>		

## Report:

The chloroplast ATP synthase, a complex multi-subunit enzyme of the thylakoid membrane, catalyses ATP synthesis and ATP hydrolysis coupled with a transmembrane proton transport. The enzyme consists of 2 distinct structural domains. The membrane integrated CF<sub>o</sub>-complex mediates proton transport across the thylakoid membrane and provides specific sites for the attachment of the membrane extrinsic, catalytic CF<sub>1</sub>-complex which contains five different polypeptides in a stoichiometry of  $\alpha_3\beta_3\gamma\delta\epsilon$ . The chloroplast ATP synthase shares many structural and functional characteristics with the homologous bacterial and mitochondrial ATPases, but is unique in several aspects of enzyme activation and sensitivity towards specific energy transfer inhibitors [1].

In order to understand these structural and functional differences we study the chloroplast ATPase from spinach and various inhibitor complexes of the membrane extrinsic CF<sub>1</sub>-domain.

We have recently solved the structure of a  $\alpha_3\beta_3$ -core complex from spinach chloroplast ATPase [2] by molecular replacement using atomic coordinates derived from the homologous bovine mitochondrial F<sub>1</sub> [3]. The structure probably reflects a unique latent state of the chloroplast enzyme which is not found in bacterial and mitochondrial ATPases. The chloroplast  $\gamma$  and  $\epsilon$  subunits which control the activation of the CF<sub>1</sub>-complex were not resolved in this native structure.

In order to obtain additional structural information on these subunits and to analyse the mechanism and the binding sites of certain energy transfer inhibitors we have started to study various CF<sub>1</sub>-inhibitor complexes.

Data sets of CF<sub>1</sub>-LuciferYellow, CF<sub>1</sub>-OPDM and CF<sub>1</sub>-Quinacrine were collected during 3 shifts on ID14-2. Data were processed by DENZO/SCALEPACK on site. All crystals showed high mosaicity, belong to the space group *R32* and have unit cell parameter similar to the native CF<sub>1</sub>-complex.

CF <sub>1</sub> -LuciferYellow	3.5 Å, 12.1 % R-merge, 93.3 % completeness, 2.7 I/σI (outer shell)
CF <sub>1</sub> -OPDM	3.5 Å, 13.3 % R-merge, 96.2 % completeness, 2.0 I/σI (outer shell)
CF <sub>1</sub> -Quinacrine	3.6 Å, 11.0 % R-merge, 98.5 % completeness, 3.0 I/σI (outer shell)

Refinement of the CF<sub>1</sub>-inhibitor complexes is in progress. In addition to the data sets collected on the various inhibitor-complexes diffraction of several CF<sub>1</sub>-tentoxin crystals was tested in order to optimise cryo-conditions.

## References:

- [1] Groth, G. und Strotmann, H. (1999) *Physiologia Plantarum* 106: 142-148.
- [2] Groth, G. and Pohl, E. (2001) *J. Biol. Chem.* 276, 1345-1352
- [3] Abrahams, J.P., Leslie, A.G.W., Lutter, R. and Walker, J.E. (1994) *Nature* 370, 621-628