



FIP -BM30A: **an automated beamline** **for protein crystallography**

Automated data collection (Xnemo)

Cryogenic Automated Transfer System (CATS)

Automated Data Processing (ADP)



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The FIP beamline

- installed on a Bending Magnet section (BM30A) at ESRF
- devoted only for protein crystallography
- tunable energy for anomalous scattering use (MAD/SAD)
- financed by:

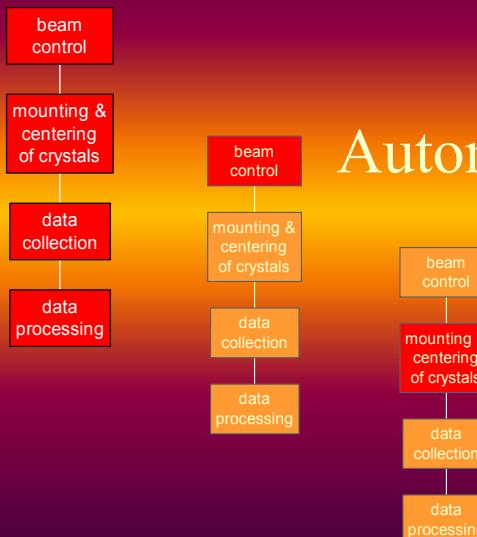


Beam time

- 33% ESRF users
- 7 % Belgium users
- 60 % French users



Automation of the crystallography experiment



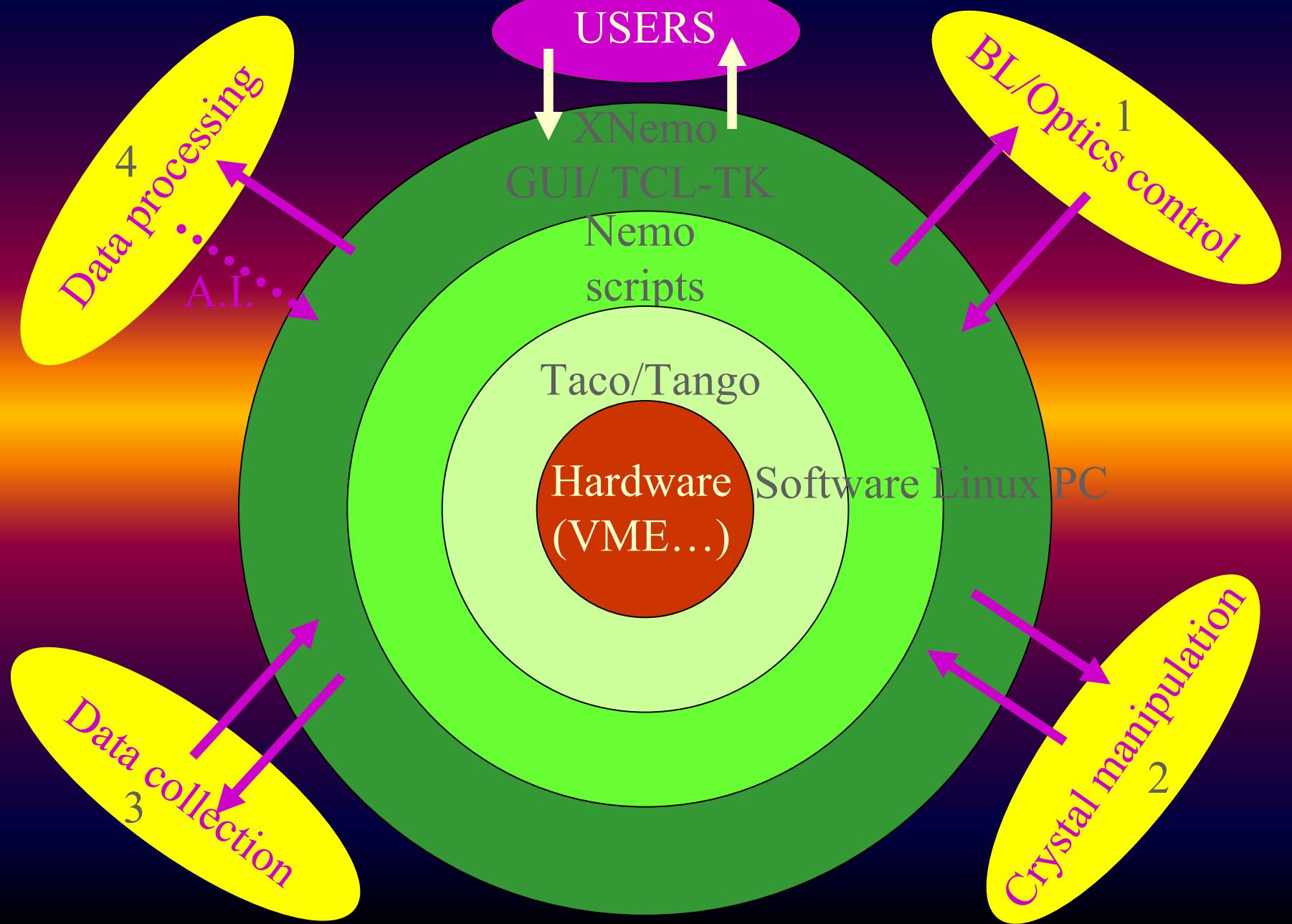
Automation of the beam control (1998)

Automation of mounting and centering (2003)

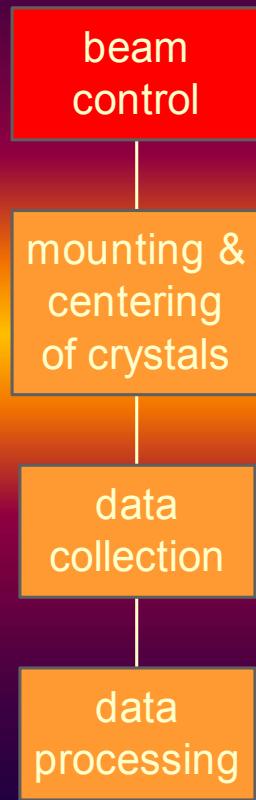
Automation of data collection (1999)

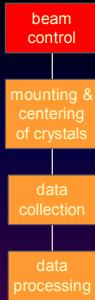
Automation of data processing (2001)

Architecture for experiments management

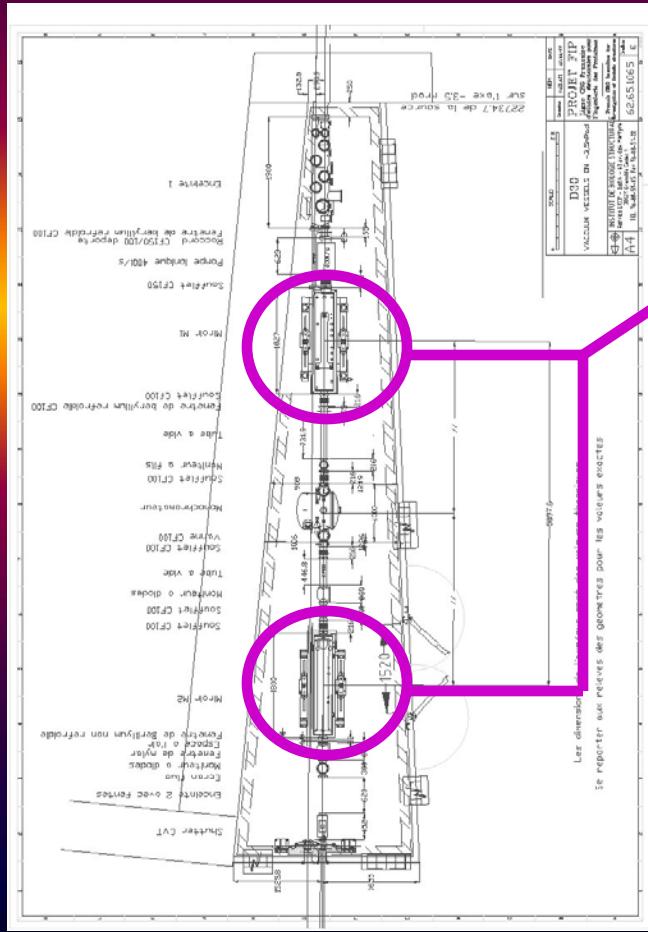


1- Automated experiment: beam control



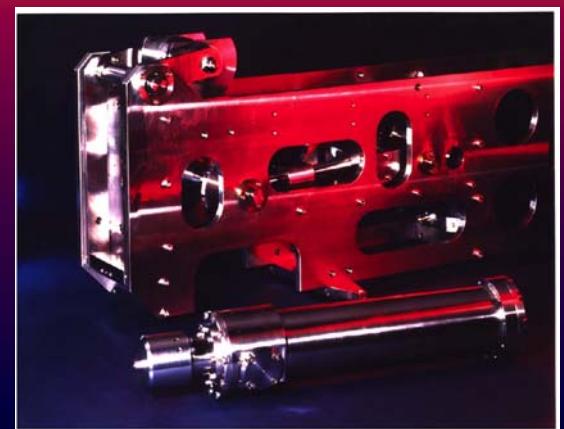


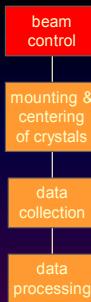
Two grazing angle mirrors



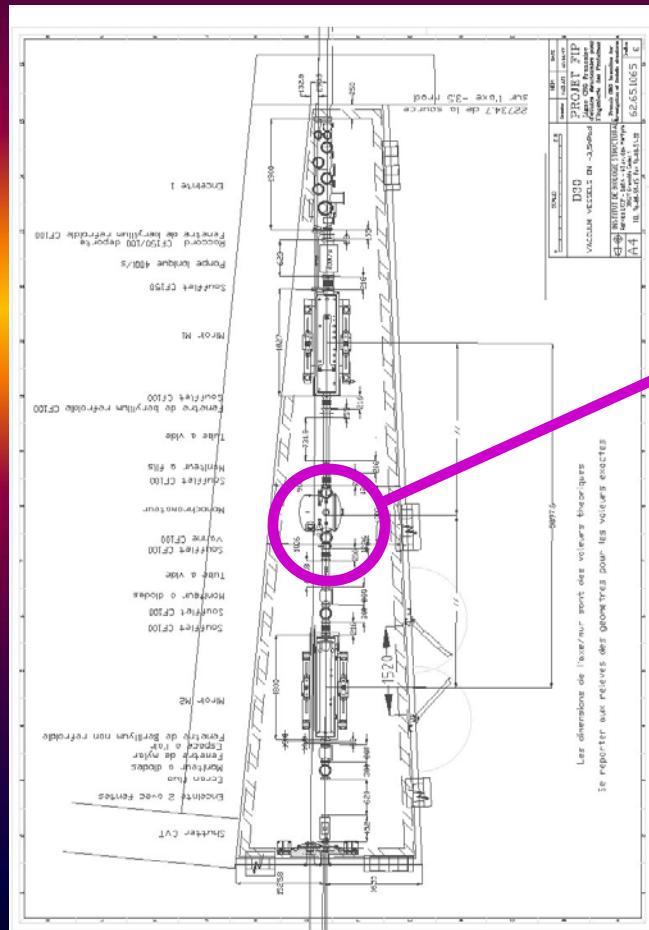
1.3 m long, 6 cm large
Si / Zerodur™
water cooling (Ga bath)

vertical focusing
harmonics rejection
heat load reduction

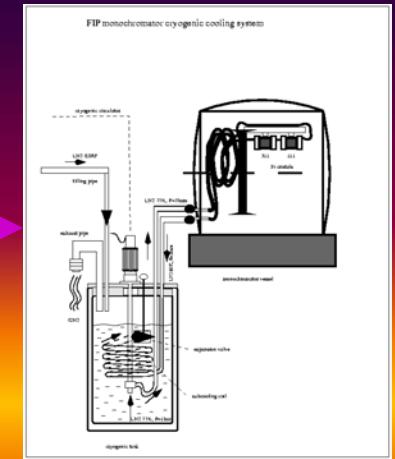
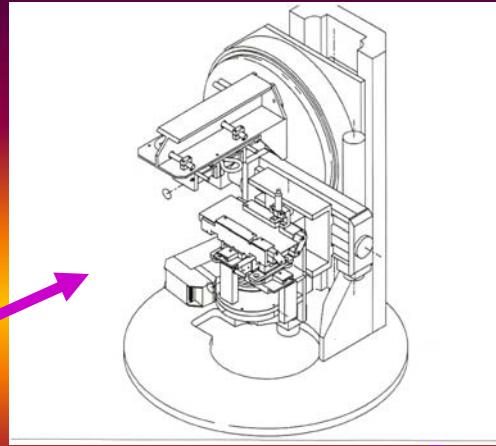




Two crystals monochromator (13 motors!)



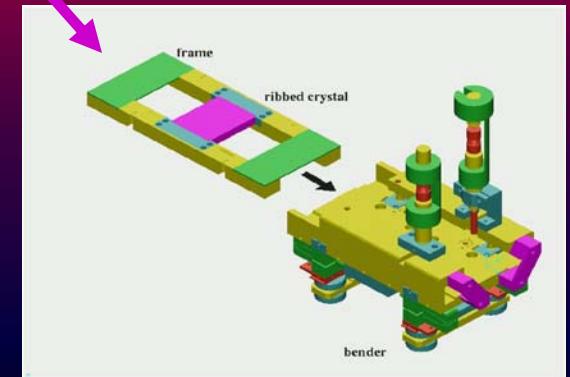
$2\text{Si}(111)$



heat load reduction

Monochromatic:
 $7 \text{ keV} < E < 17 \text{ keV}$
 $\Delta E/E \sim 10^{-4}$

Focal spot:
 $300*300 \mu\text{m}^2$

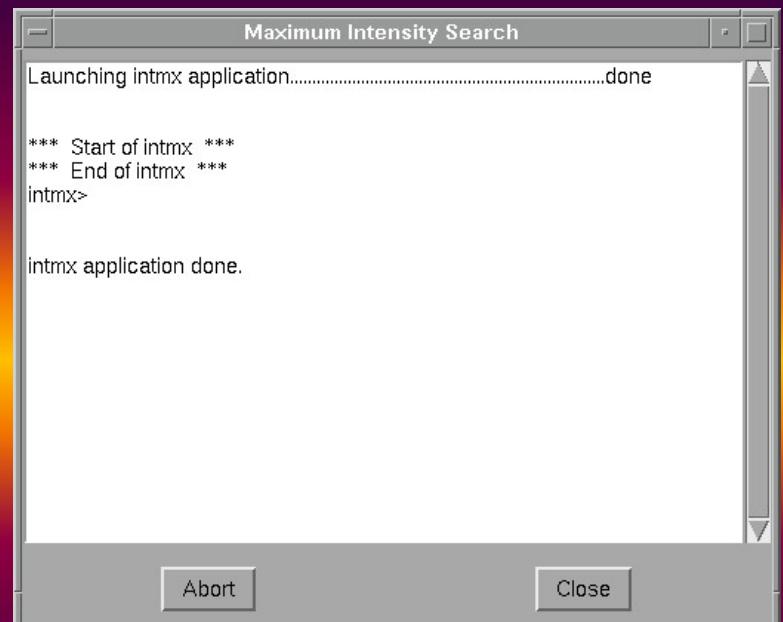
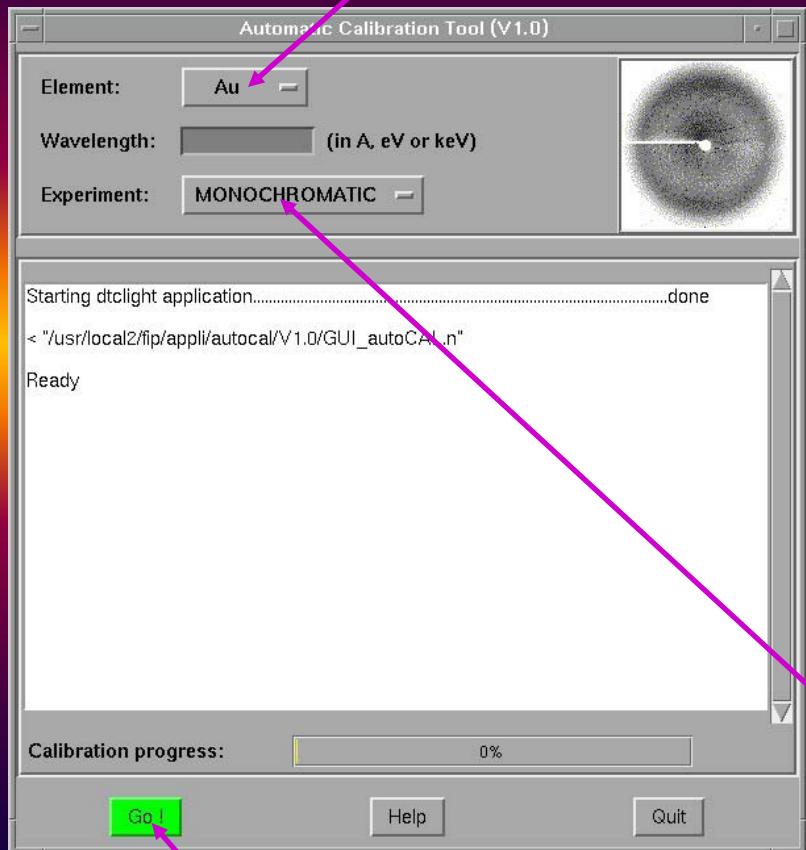


horizontal focusing



Setting up the X-ray beam

1. select edge or wavelength



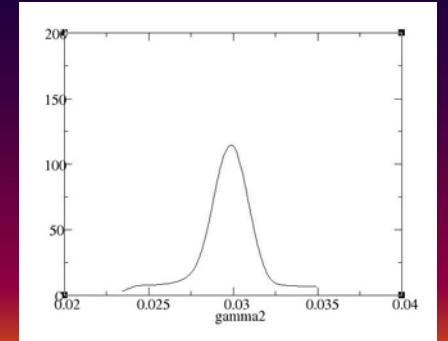
2. theoretical position, or automated calibration with metal foil

3. and then: Go...

Setting up the X-ray beam

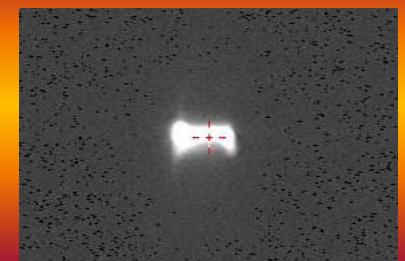
- optics geometry (theory)

- * based on absolute position of origin
- * tabulated corrections

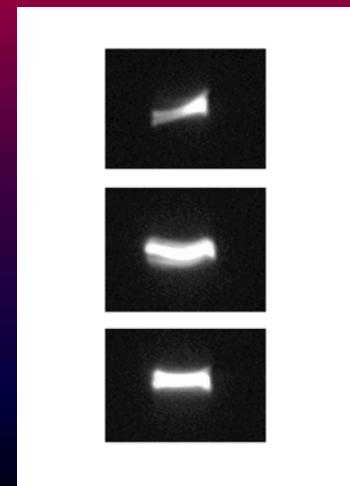
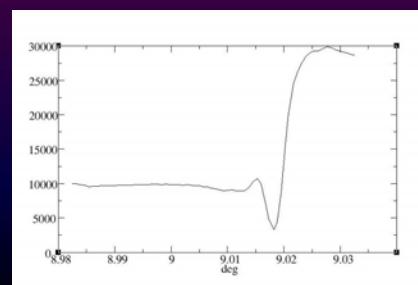


- beam optimization

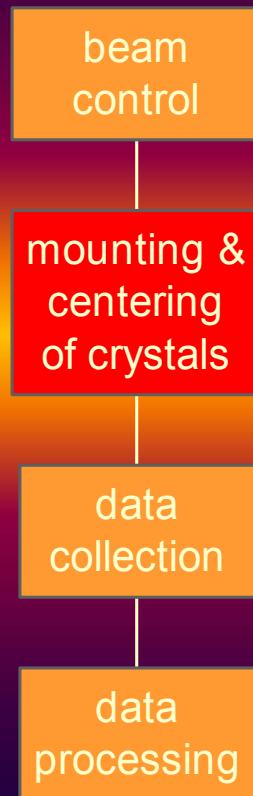
- * scans (before collimator)
- * beam position on fluo.
- * scans (after collimator)
- * beam shape on fluo.



- energy calibration (optional)

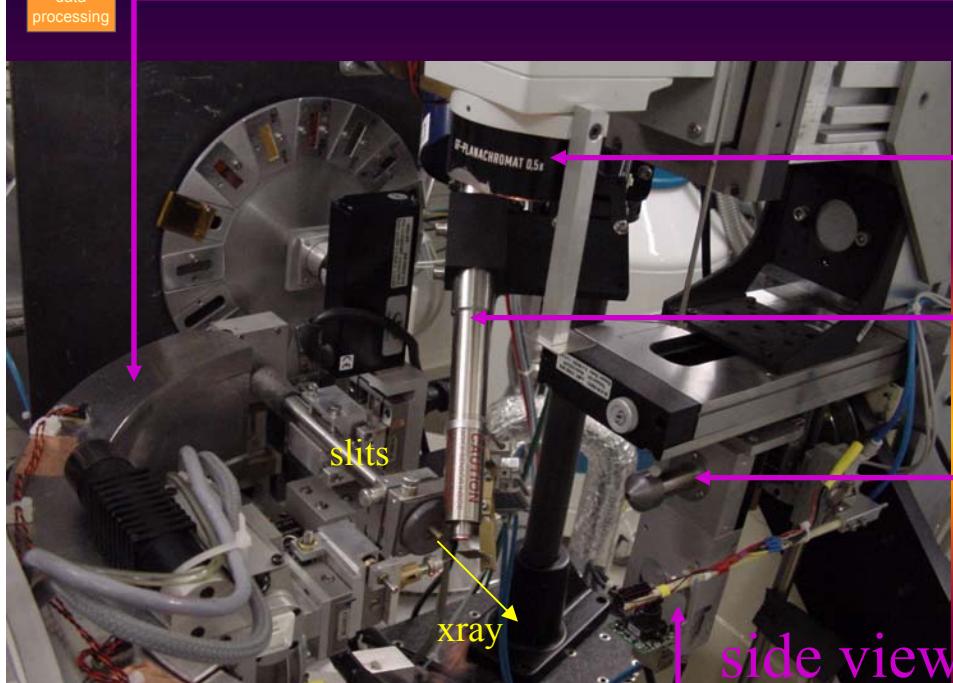
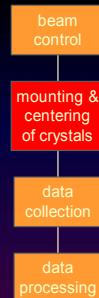


2- Automated experiment: sample mounting and centering



The experiment

A 5-circle diffractometer including:



- beam monitors

- 2-theta for high resolution

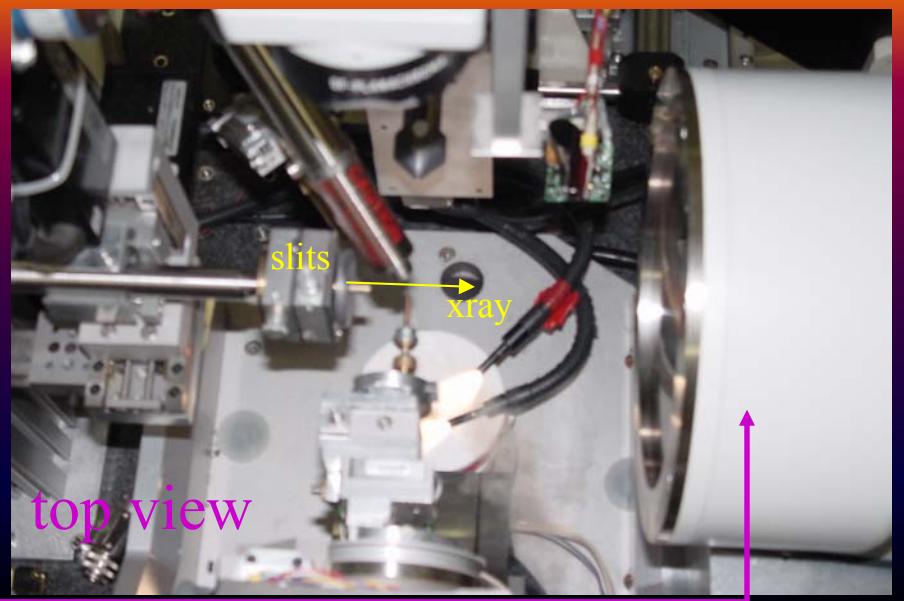
- Mar CCD detector

- sample orientation (MAD)

- microscope + video

- Oxford cryo. cooling

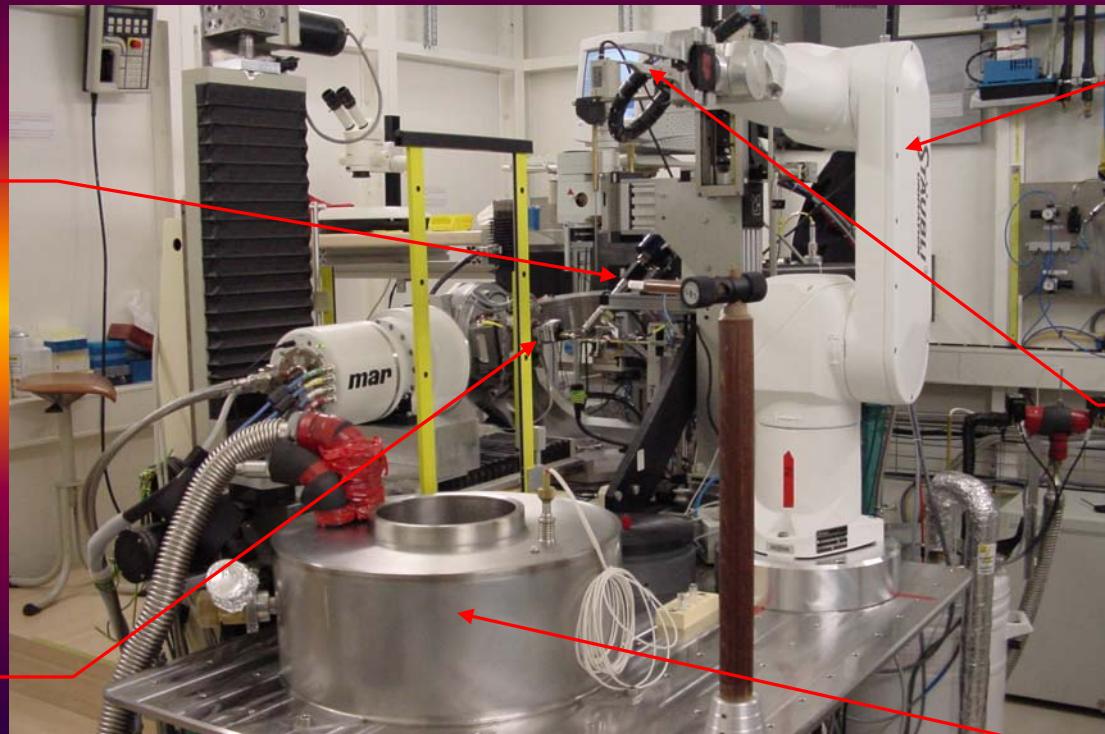
- fluorescence detector





Integrated system

Flipping tong



Robot

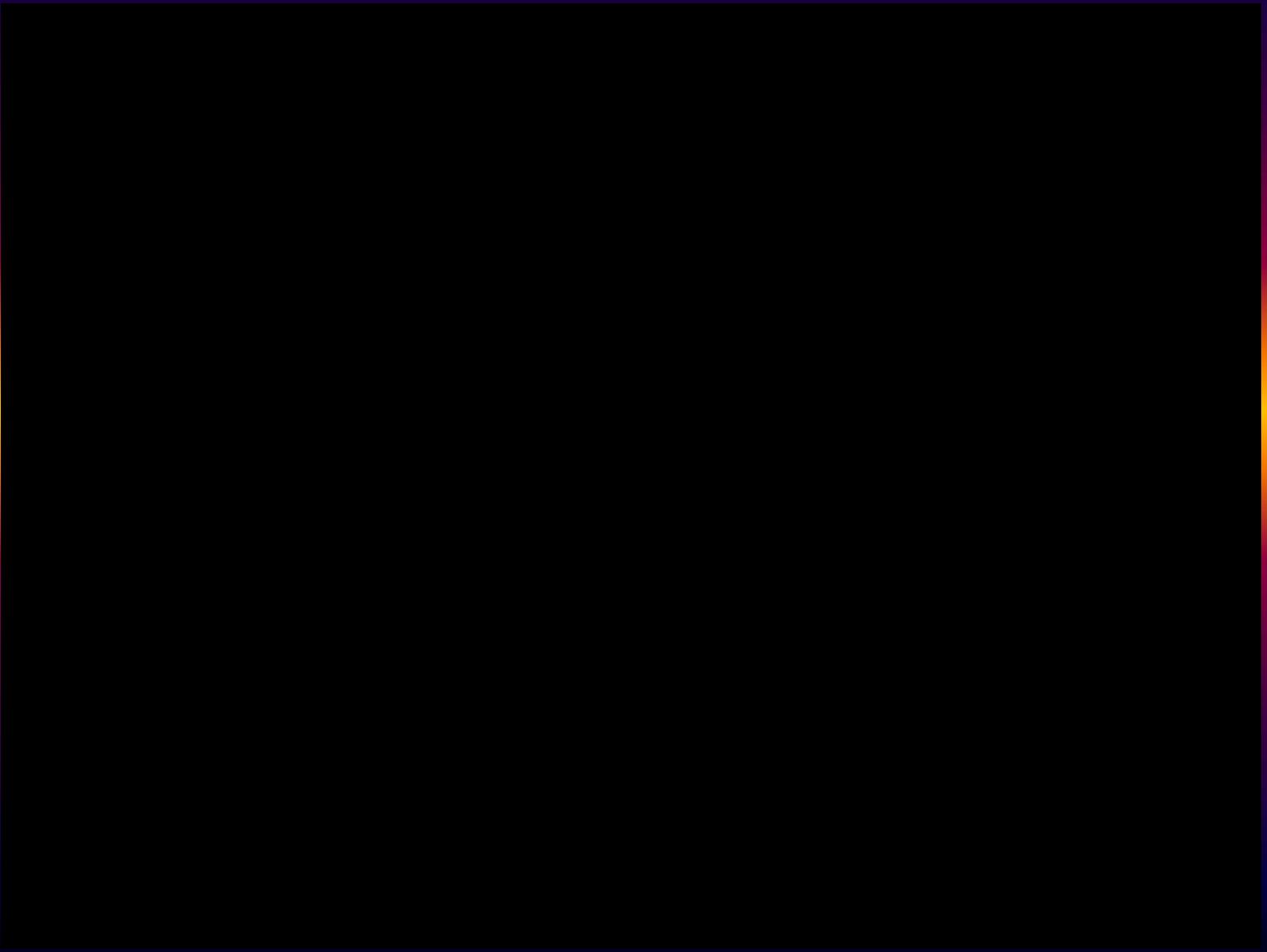
Actuator

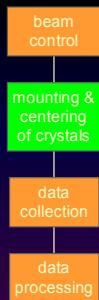
Goniometer
head

Storage
Dewar

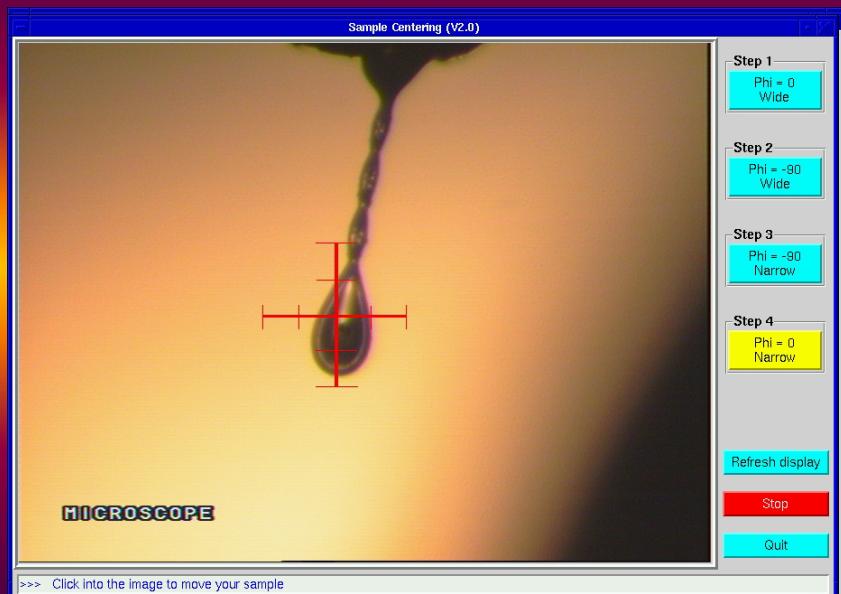


Sample unmounting (magnetic cap)

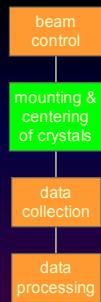




Semi-automated crystal centering



... steps 1, 2, 3 and 4
in chronological order



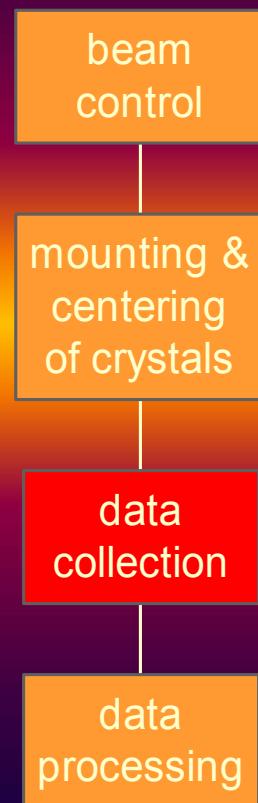
Automated crystal centering

Crystal recognition
standard

UV

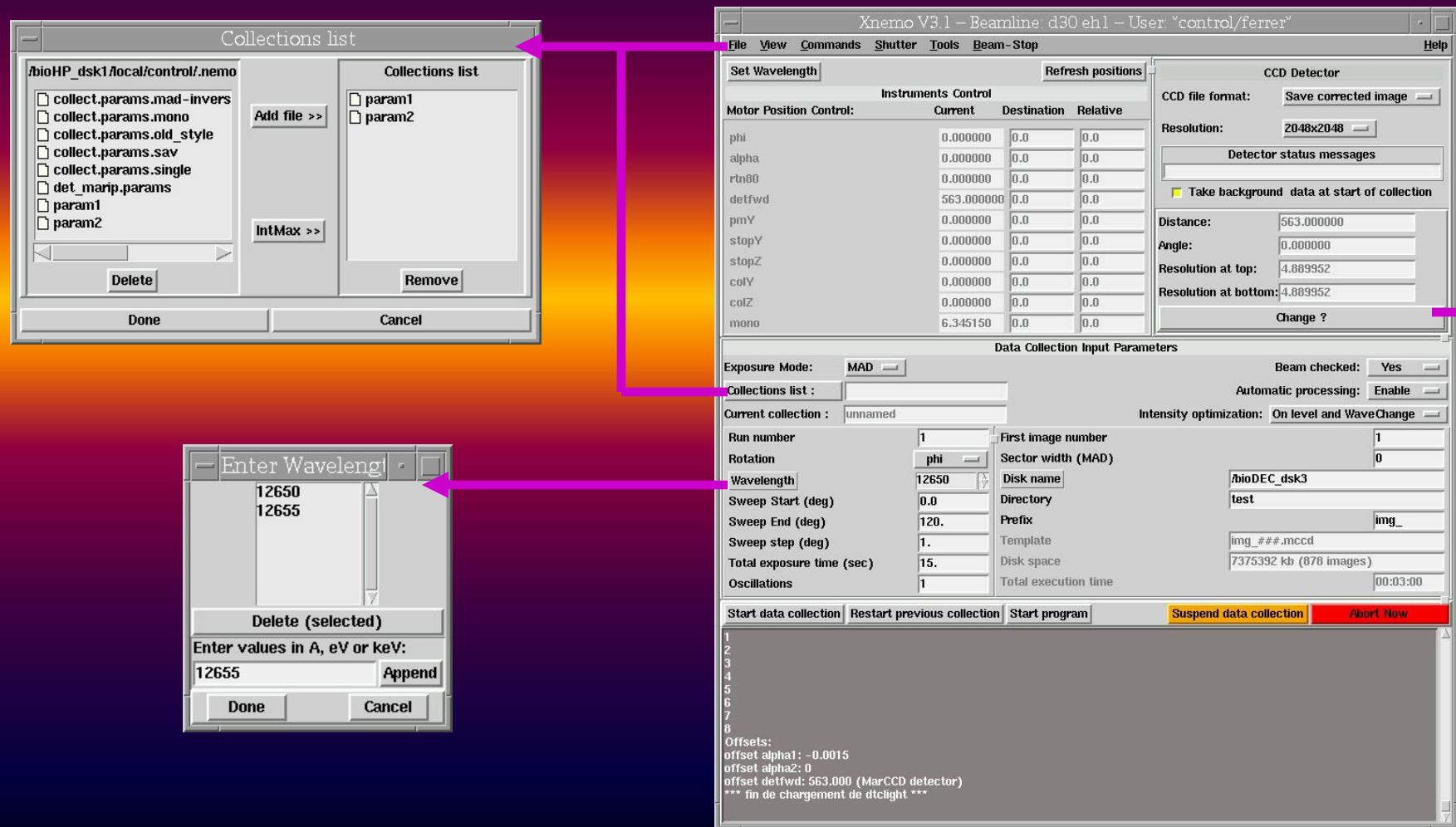


3-Automated experiment: data collection





Preparing data collection



Xnemo

Workflow Diagram:

```

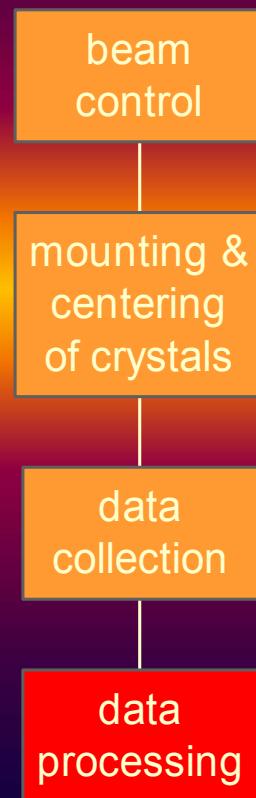
graph TD
    A[beam control] --> B[mounting & centering of crystals]
    B --> C[data collection]
    C --> D[data processing]
  
```

Xnemo V3.1 - Beamline: d30 eh1 - User: "control/ferrer"

Software Components and Data Flow:

- Beamline Control:** Xnemo V3.1 interface showing motor position control (phi, alpha, rtm60, detfwd, pmY, stopY, stopZ, colY, colZ, mono) and data collection input parameters.
- Mounting & Centering:** CCD Detector configuration window (CCD file format: Save corrected image, Resolution: 2048x2048).
- Data Collection:** Beam checked: Yes, Automatic processing: Enable, Intensity optimization: On level and WaveChange.
- Data Processing:** Data collected on FIP (BM30A) shown in a Netscape browser window.
- Analysis:** A second Netscape browser window displays the dataset collected on FIP (BM30A) and a third window shows the raw data log file (img_001.log) containing command history and log entries.

4- Automated experiment: data processing





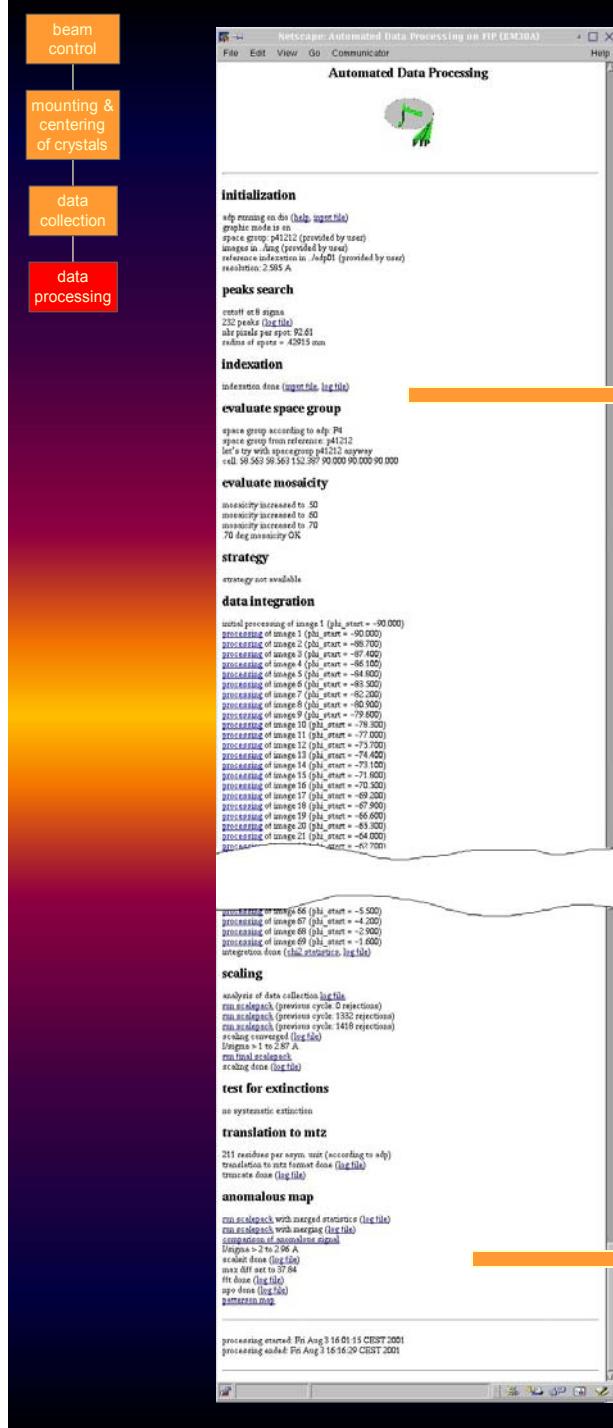
adp: steps

peak search (MarSearch)
indexation (denzo)
mosaicity
crystal reorientation (oXo)
strategy (strategy/best)
integration (denzo)
partials
scaling (scalepack)
extinctions
Patterson map, ... (CCP4)

⇒ peak size ⇒ box size
⇒ laue group, nbr. residues
⇐ *denzo log file (iterations)*
⇒ expected completeness
⇐ *interruptions*
⇒ anomalous signal
⇒ final space group
⇒ max resol. for anomalous

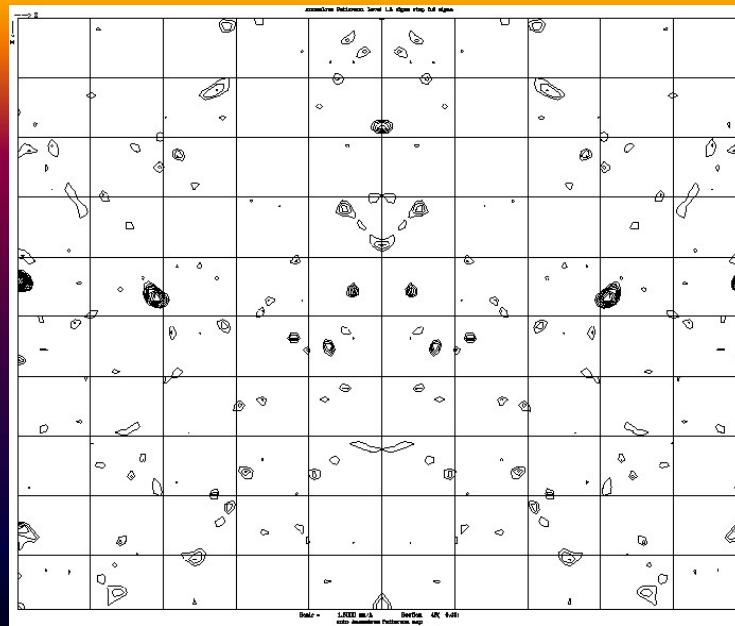
site search (solve) molecular replacement (molrep)
heavy atoms
SAD phasing (solve)
solvent fl. (resolve)
model building (resolve)

adp: log file



This screenshot shows the continuation of the log file content from the previous window.

Lattice	metric tensor	best cell (uncentered)	best cell (without symmetry constraints)
primitive cubic	19.94% 115.66 109.04 101.62 117.26 121.04 59.11	111.50 111.50 111.50 111.50 111.50 111.50 30.00	50.00 30.00
I centred cubic	6.44% 117.24 115.66 117.24 117.24 117.24 117.24 59.55	59.33 59.64	59.55 59.64
F centred cubic	3.10% 115.66 113.71 117.14 117.14 117.14 117.14 59.25	59.25 59.25	59.25 59.25
primitive rhombohedral	3.09% 115.66 109.04 109.02 64.74 59.36 53.51	111.50 111.50 111.50 111.50 111.50 111.50 31.07	51.07 51.07
primitive hexagonal	15.07% 109.02 111.45 112.54 97.59 64.74 59.95	110.46 111.45 112.54 97.59 64.74 59.95	64.74 59.95
primitive tetragonal	19.94% 109.92 111.65 109.04 120.43 64.74 121.04	112.74 112.74 112.74 112.74 112.74 112.74 30.00	50.00 30.00
I centred tetragonal	0.78% 117.17 115.66 147.14 117.14 117.14 117.14 59.53	59.53 59.64	59.53 59.64
primitive orthorhombic	19.94% 109.04 109.02 115.66 115.66 115.66 115.66 59.28	111.50 111.50 111.50 111.50 111.50 111.50 30.00	59.28 59.28
C centred orthorhombic	15.10% 109.02 110.41 109.04 110.31 64.74 52.73	109.02 110.41 109.04 110.31 64.74 52.73	50.00 30.00
I centred orthorhombic	0.65% 115.66 117.17 147.14 117.14 117.14 117.14 59.26	115.66 117.17 147.14 117.14 117.14 117.14 30.00	50.00 30.00
F centred orthorhombic	0.67% 117.24 113.71 116.55 116.55 116.55 116.55 59.75	59.75 59.84	59.75 59.84
primitive monoclinic	15.28% 109.92 110.04 111.07 111.07 111.07 111.07 59.24	110.92 110.92 110.92 110.92 110.92 110.92 31.24	59.24 59.24
C centred monoclinic	0.27% 104.85 117.17 115.66 90.64 117.28 93.55	104.85 117.17 115.66 90.64 117.28 93.55	117.28 93.55
primitive triclinic	0.00% 103.04 103.82 111.07 116.86 96.11 115.26	103.04 103.82 111.07 116.86 96.11 115.26	96.11 115.26
centroids unit cell	117.16 117.16 147.09 90.00 90.00 90.00	90.00 90.00	90.00 90.00
crystal rotz, roty, rotx	72.933 -141.928 -40.530		
Autotindex ZB=em, T3em	99.40 -82.71		

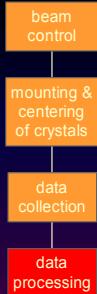




3.8 Å dataset processed by user vs adp

I	sigI	Average stat.	Norm.	Linear	Square				
			Chi**2	R-fac	R-fac				
3356.8	285.7	267.0	0.451	<u>0.125</u>	0.083	◀	user		
3281.8	259.9	238.4	0.552	<u>0.128</u>	0.090	◀	adp		
% of reflections with I / Sigma less than									
0	1	2	3	5	10	20	>20	total	
6.9	23.1	34.4	42.5	53.6	69.3	84.1	15.4	<u>99.5</u> ▶	user
6.1	21.4	32.6	40.5	51.5	68.0	83.4	16.2	<u>99.6</u> ▶	adp

Comparison of a typical dataset, processed by a crystallographer using *HKL* and with *adp*.
 Each data processing is illustrated with statistics calculated by *scalepack*
 (upper part: I, K^2 and R-factors; lower part: I/ $\sigma(I)$ and completeness).



First structure solved in automated mode

Structure solved in
3h data collection
2h phasing/building

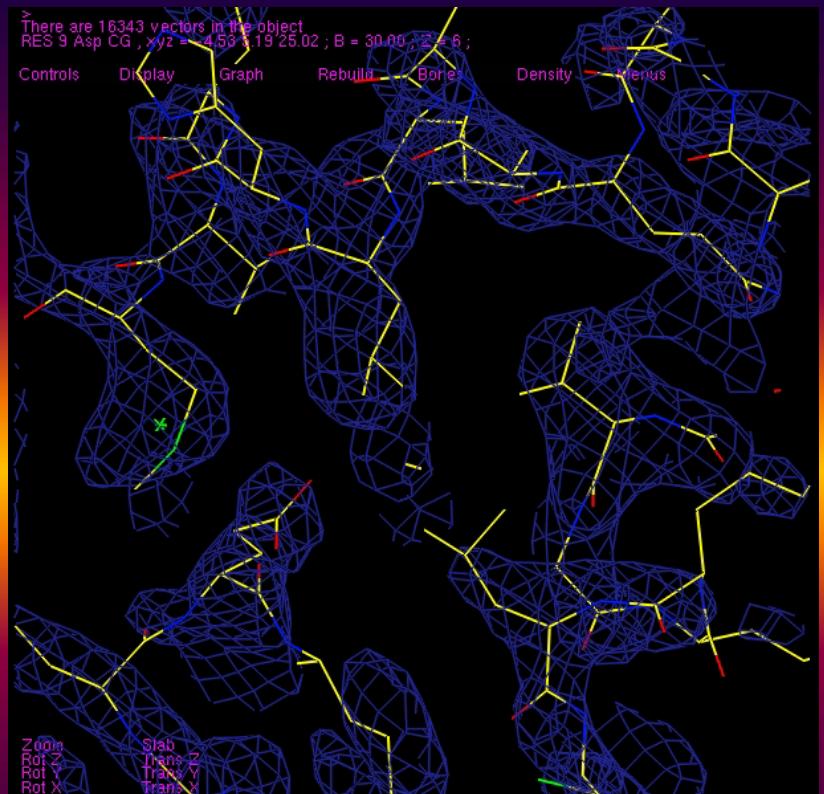
Data collection statistics:

p212121, 1 mol./asym. unit
completeness at 2.1 \AA : 98.7%
 $\langle R_{\text{sym}} \rangle$: 4.2%
 R_{sym} (last shell) : 12.5%

Phasing statistics (solve/resolve):

fig. of merit: 0.25
180/233 residues built autom.

80% of the model in 5 hours



Farnesoid X Receptor (FXR),
M. Downes et al.,
Molecular Cell 11 (2003), 1079



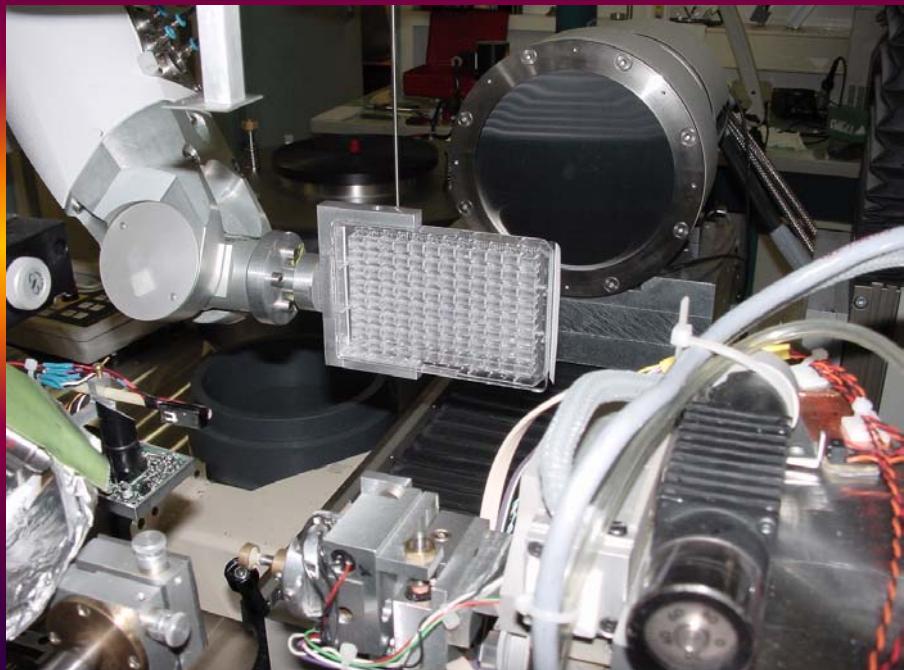
Analysis of crystallization drops

Aim:

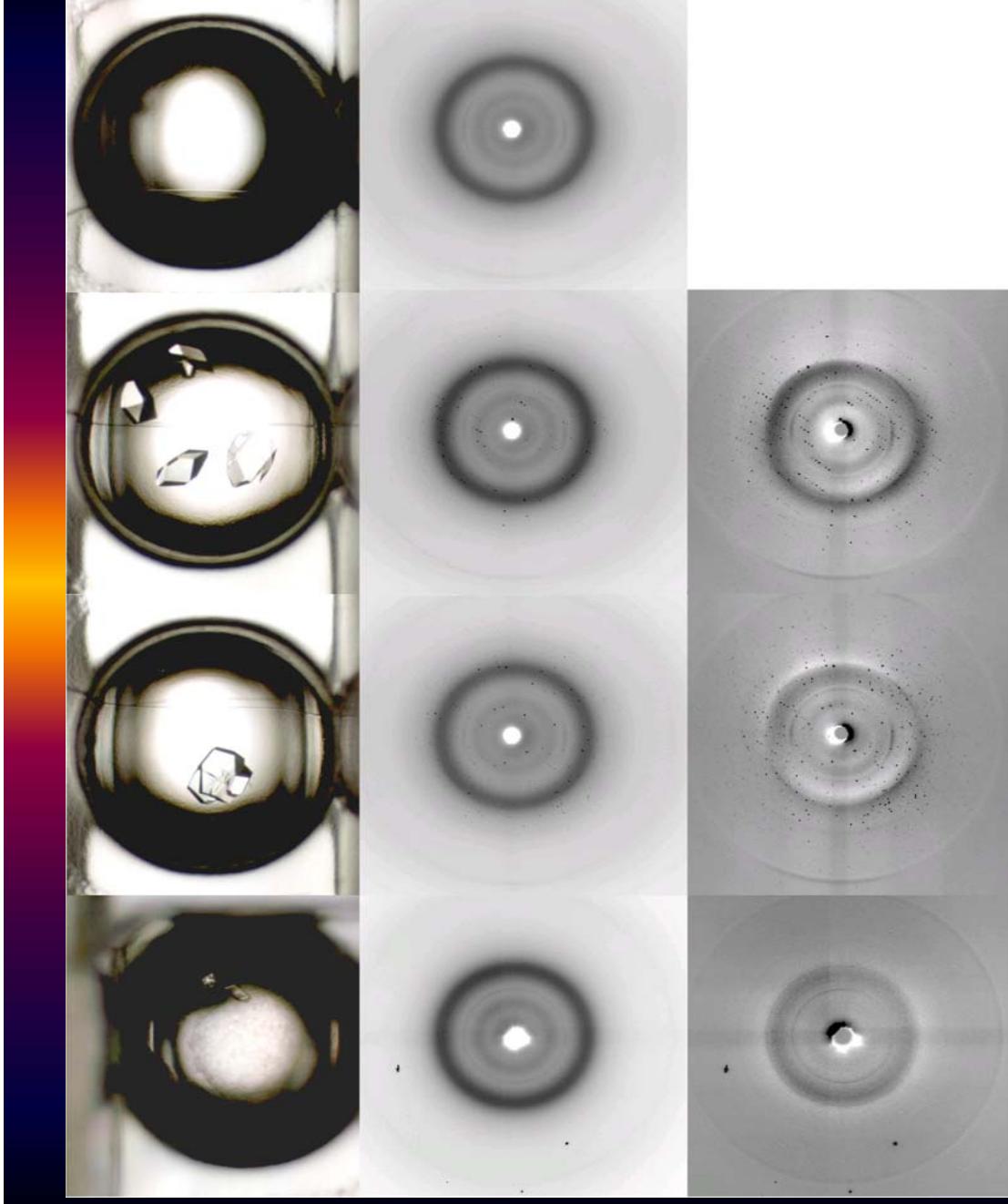
automated analysis of Greiner™ box
discrimination salt/protein
precipitate analysis ?

Means:

beam: 2x2 mm, 0.8 Å
oscillations: 1 deg
10 to 30 sec / drop (1 to 2 h per box)
image processing



Protein *vs.* Salt



← Protein crystal

← Salt crystal

Precipitate analysis !

Data collection *in situ*

<i>Plate-drop</i>	L-B3b	T-C12a	T-B4b	C-B1c	Y-A11a
<i>sample</i>	lysozyme	lysozyme	thaumatin	chalcone s.	kinase
<i>Space group</i>	p43212	p43212	p41212	p3121	c2
<i>Resolution (Å)</i>	1,8	1,8	2,2	3,0	2,2
<i>Completeness (%)</i>	89,8	85,1	95,1	70,0	48,2
<i>Redundancy</i>	2,6	2,8	2,4	2,6	1,3
<i>I/σ</i>	11,0	9,1	6,1	4,2	6,1
<i>Rsym (%)</i>	6,2	8,4	11,5	23,9	7,1
<i>Rfree (%)</i>	26,9	26,7	27,2	24,2	28,9

L. Jacquemet et al., accepted in Structure

J.-L. Ferrer	(beamline responsible)
L. Jacquemet	(crystallographer)
J. Joly	(software/hardware)
P. Charrault	(electronic)
M. Pirocchi	(vacuum)
J. Ohana	(robotics)
R. Kahn	(adviser)
F. Borel	(local contact)
S. Fieulaine	(local contact)
L. Serre	(local contact)
J. Dupuis	(local contact)
P. Israel-Gouy	(administration)
A. Bertoni	(machining)
M. Roth (retired)	(adviser)
P. Carpentier (IBS/BM16)	(instrumentation)
E. Fanchon (IBS/LCM)	(software)