

## Introduction

The goal of the BioSAXS experiment with number MX-1912 was to study the protein KAP1 in solution. KAP1 or Krüppel associated protein 1, is a multidomain protein that has been linked to the development and differentiation of many adult cell types as well as many other fundamental cellular processes including gene silencing, transcription regulation and DNA damage repair [1, 2]. It acts like a scaffold protein recruiting many different proteins and enzymes to influence the organization of chromatin structure. Because of that, we are interested in finding out the three dimensional structure of KAP1 and understanding the molecular interactions between KAP1 and the proteins it recruits.

## Data acquisition

The data were recorded at the ESRF BM29 over a  $q$  range of  $0.025$ - $5 \text{ nm}^{-1}$  and beam wavelength of  $0.992 \text{ \AA}$ . Protein samples were exposed under flow through a capillary at a rate of  $10$  or  $8 \text{ \mu L}\cdot\text{s}^{-1}$  with  $10$  individual frames collected and averaged to give the final scattering profile. The first measurements clearly indicated that buffer containing high salt and glycerol stabilized the protein and protected it from radiation damage, so all subsequent measurements were done in that buffer. The dilution series was done diluting the protein in buffer (a). The blank for each measurement was the corresponding flow through from protein concentration. Scattering from this blank alone was collected before and after each sample to correct for fluctuations in beam intensity and to clean the capillary between samples. The temperature was always  $20^\circ \text{ C}$  and the viscosity was set to low. We used  $100\%$  of the beam transmission. Four variants of KAP1 of different lengths were measured: The concentration series ranged from  $7$  to  $0.1 \text{ mg/ml}$ .

## Data analysis

Images were analyzed using the ATSAS package [3]. PRIMUS [4] and GNOM [5] were used to obtain Guinier plots and compute pair-distribution functions, respectively. Scattering curves for the different concentrations coincided for most of the measured  $q$  range but showed signs of aggregation at very low  $q$  values.

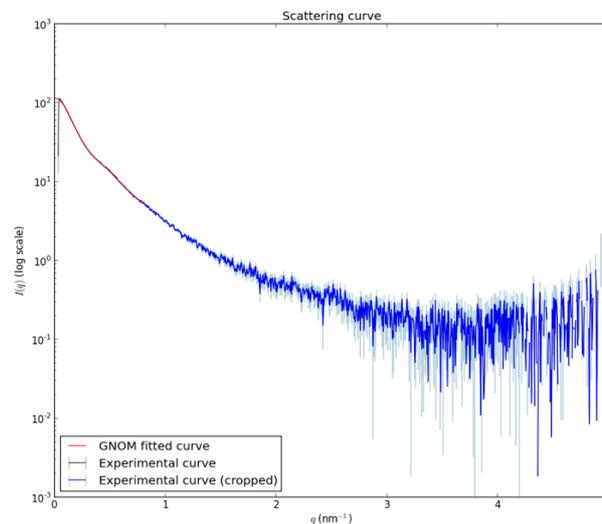


Figure 1: Typical scattering curve showing signs of aggregation at low  $q$  values.

## Results and Conclusions

Preliminary analysis of the data collected shows that KAP1 is elongated. Structural information obtained by SAXS, in combination with our ultracentrifugation and light scattering data, would be very useful to validate the 3D models and assess the sample

heterogeneity and suitability to perform further crystallographic and electron microscopy experiments. The next step is to apply for more beam time to be able to perform size exclusion chromatography (SEC) before the SAXS measurement, to perhaps improve the measurement at low  $q$  values. Please see the application for the next shift.

## References

1. Iyengar, S. and P.J. Farnham, *KAPI protein: an enigmatic master regulator of the genome*. J Biol Chem, 2011. **286**(30): p. 26267-76.
2. Cheng, C.T., C.Y. Kuo, and D.K. Ann, *KAPtain in charge of multiple missions: Emerging roles of KAPI*. World J Biol Chem, 2014. **5**(3): p. 308-20.
3. Petoukhov, M.V., et al., *New developments in the ATSAS program package for small-angle scattering data analysis*. J Appl Crystallogr, 2012. **45**(Pt 2): p. 342-350.
4. Konarev, P.V., et al., *PRIMUS: a Windows PC-based system for small-angle scattering data analysis*. Journal of Applied Crystallography, 2003. **36**: p. 1277-1282.
5. Svergun, D.I., *Determination of the Regularization Parameter in Indirect-Transform Methods Using Perceptual Criteria*. Journal of Applied Crystallography, 1992. **25**: p. 495-503.